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DATA SHEET

Version: 03
Revision date: 12/04/2023

1. Identification

Product name **Clean-Easy Agarose Purification Kit**

Cat. No **AN0070 (50 reactions)**
Cat. No **AN0071 (100 reactions)**
Cat. No **AN0071-XL (250 reactions)**

2. Description

Clean-Easy Agarose Purification Kit provides a rapid and efficient method to extract DNA from agarose gels. It is based on the solubilisation and binding of DNA to a silica membrane in presence of chaotropic salts. Clean-Easy minispin columns contains an exclusive membrane that allows to bind an unique DNA fragment, previously excised from agarose gel.

3. Kit Components

Item	Quantity		
	AN0070 (50 rxn)	AN0071 (100 rxn)	AN0071-XL (250 rxn)
Clean-Easy minispin columns	50	100	250
Collection tubes (2 mL)	100	200	500
QG Buffer	60 ml	100 ml	200 ml
PE Buffer*	11.25 ml	22.5 ml	45 ml
EB Buffer	20 ml	40 ml	100 ml

* Add ethanol (96%-100%) [not included] to PE Buffer prior to initial use. After ethanol has been added, mark the bottle to indicate that this step has been completed.

4. Features

- **Simple** and **Just a few minutes** procedure.
- **Wide spectrum of size** fragments could be purified, (suitable since 100 bp up)
- **High Percentage of Recovery**, greater than 80% on 0.7-1% agarose. Recovery is lower in more concentrated agarose gels (50-60% on 2% agarose).
- DNA purified **Ready to use** in all molecular biology procedures.
- Suitable for any kind of agarose and gel buffer systems.

5. Storage specifications

Clean-Easy Agarose Purification Kit should be stored at room temperature (15–25°C) for up to 12 months without any reduction in performance.

6. Applications

- Purification of DNA fragments (*obtained by PCR or digestion with restriction enzymes)* from agarose gels.
- The purified DNA can be used in all molecular biology applications.

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7. Quality Certifications

Clean-Easy Agarose Purification Kit is tested in the purification of a 0.5 kb DNA fragment excised from 2% agarose gel. The purified band is analysed in agarose gel electrophoresis.

8. Further information

Product Use Limitations

This product is developed, designed, and sold exclusively only for research purposes use. The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals.

Safety Information

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, please consult the appropriate material safety data sheets (MSDS). These are available online in convenient and compact PDF format at www.canvaxbio.com where you can find, view, and print the MSDS for each CANVAX kit.

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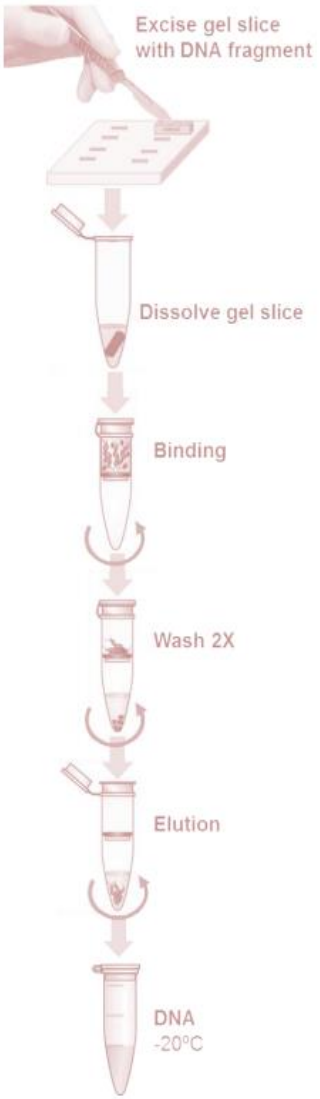
DETAILED PROTOCOL

1. Using a clean, sharp razor blade or scalpel, excise the DNA ban from the agarose gel. Remove the extra agarose to reduce the size of gel slice. Place the gel slice in a 1.5 ml preweighted tube and weigh the gel slice (The maximum amount of gel slice per columna is 400 mg).



2. Add 3 volumes of QG Buffer to 1 volume of gel. (For example, if the agarose gel slice is 100 mg, add 300 µl of QG buffer)

For gels containing more than 2% agarose, add 6 volumes de QG Buffer per mg of gel.

3. Incubate at 50 ˚ C in a water bath for 10 min or until the gel slice has completely dissolved. During incubation at 50 °C, mix by vortexing or inverting the tubes every 1 minute.

Make sure the gel slice completely dissolved. For >2% gels, increase incubation time.

Important! For fragments <500 bp and >4 kb, add 1 volume of isopropanol to the sample and mix (For example, if the agarose gel slice is 100 mg, add 100 µl isopropanol)

4. Label the lid of a new minispin column placed in a 2 ml collection tube. Carefully apply the mix from previous step to the spin column and Centrifuge at 13000 rpm for 1 minute. For mixture volumes of more than 750 µl, load and centrifuge again using the same column.

5. Place the spin column in a new 2 ml collection tube, and discard the collection tube containing the filtrate. Add 700 µl of PE buffer to the minispin column and centrifuge at 13000 rpm for 1 minute.

Remember! Before using for the first time, add ethanol (96–100%) to PE Buffer as indicated on the bottle.

6. Discard the flow-through and centrifuge again at 13000 rpm for 1 minute. This step is essential for removing trace buffer PE.

7. Transfer the column to a clean 1.5 ml microcentrifuge tube. Add 30 µl of Elution Buffer (EB) or H2O (pH=7.0-8.5) to the center of the column membrane and incubate at room temperature for 1 minute. Centrifuge at 13000 rpm for 1 minute to elute and collect DNA.

*To increase the DNA yield you can warm the buffer EB/H2O to 65 ˚C